

Dear author,

Please note that changes made in the online proofing system will be added to the article before publication but are not reflected in this PDF.

We also ask that this file not be used for submitting corrections.

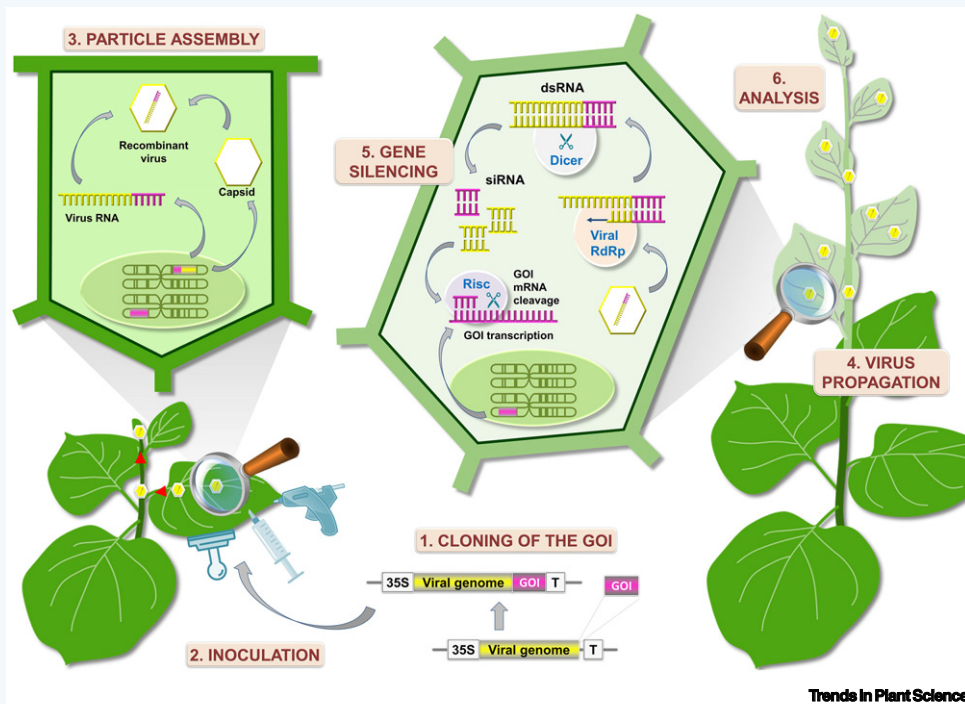
# Virus-Induced Gene Silencing: Hush Genes to Make Them Talk

Vincent Courdavault,<sup>1,\*</sup> Sébastien Besseau,<sup>1</sup> Audrey Oudin,<sup>1</sup> Nicolas Papon,<sup>2</sup> and Sarah Ellen O'Connor<sup>3,\*</sup>

<sup>1</sup>Biomolécules et Biotechnologies Végétales, BBV, EA2106, Université de Tours, Tours, France

<sup>2</sup>Groupe d'Etude des Interactions Hôte-Pathogène, GEIHP, EA3142, Université d'Angers, SFR 4208 ICAT, Angers, France

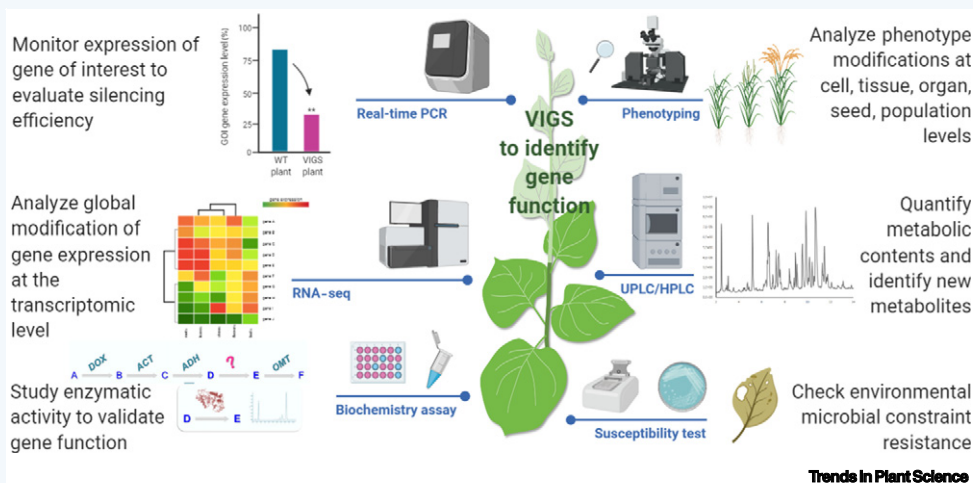
<sup>3</sup>Department of Natural Product Biosynthesis, Max Planck Institute for Chemical Ecology, Jena, Germany



Q5

Trends in Plant Science

Virus-induced gene silencing (VIGS) silences a gene of interest (GOI) by exploiting plant defense mechanisms against RNA viruses. A GOI fragment is cloned into a construct containing viral genome sequence (1) and then inoculated into plants (2), where viral particles are produced (3). Through cell-to-cell movement using plasmodesmata and systemic progression via the phloem (4), viruses infect newly developing leaves and replicate. This involves the formation of dsRNA that is cleaved by plant DICER to release siRNA. siRNAs are loaded in the multiprotein complex RISC and their antisense strand screen RNA in cells. Hybridization of this complex results in specific RNA degradation (or translation repression) of viral RNA and endogenous GOI transcripts causing its silencing.



Trends in Plant Science

After 3–6 weeks, gene silencing efficiency is analyzed by monitoring GOI expression in new aerial parts of silenced plants and plants transformed with empty vector. Gene function can be investigated by comparative studies in similar organs from both types of plant.

**ADVANTAGES:**

One of the easiest, most cost-effective, and most efficient ways to study gene function *in planta*. Allows reliable silencing of genes (up to 90%) in a short time period (routinely 2–3 weeks post-inoculation).

Wide range of procedures of virus/viral genome inoculation (e.g., agroinfiltration, agrodrench, leaf rubbing, biolistic).

Robust and straightforward protocols reported for model and non-model plants.

Allows study of essential genes that cannot be knocked out.

Large-scale VIGS experiments for EST library screening are possible.

Comparison of gene function between different species with same constructs.

May lead to DNA methylation and heritable transcriptional gene silencing.

**CHALLENGES:**

Identification of a suitable virus and inoculation method in non-model plants: virus host specificity requires tool development.

Low-level and/or nonuniform gene silencing can result in reduced phenotypic and/or metabolic alterations. Lack of observable phenotype can also result from functional redundancies between gene family members.

Symptoms caused by virus propagation may mask phenotypic variations due to gene silencing.

Gene silencing is mainly transient and does not allow long time-course studies.

Difficulties in specific silencing of individual isoforms encoded by large gene family members.

Metabolic rerouting of biosynthetic intermediates impedes identification of the reaction catalyzed by products of silenced genes.

\*Correspondence: [vincent.courdavault@univ-tours.fr](mailto:vincent.courdavault@univ-tours.fr) (V. Courdavault) and [oc Connor@ice.mpg.de](mailto:oc Connor@ice.mpg.de) (S.E. O'Connor).

Q1  
2  
Q2 Q3 Q4  
4  
5  
6  
8  
10  
11  
12  
13  
14  
15  
17  
18  
19

## Acknowledgments

We acknowledge funding from the EU Horizon 2020 research and innovation program (MIAMI project-grant agreement N°814645), the ARD2020 Biopharmaceutical Program of the Région Centre Val de Loire (BioPROPHARM and CatharSIS projects), La Ligue Contre le Cancer (Yeast4LiFE), le Studium (Consortium fellowship), and ERC 788301.

20

21

Q6

28

24

## Literature

1. Ratcliff, F. *et al.* (1997) A similarity between viral defense and gene silencing in plants. *Science* 276, 1558–1560 26
2. Kumagai, M.H. *et al.* (1995) Cytoplasmic inhibition of carotenoid biosynthesis with virus-derived RNA. *Proc. Natl. Acad. Sci. U. S. A.* 92, 1679–1683 27
3. Burch-Smith, T.M. *et al.* (2006) Efficient virus-induced gene silencing in *Arabidopsis*. *Plant Physiol.* 142, 21–27 28
4. Senthil-Kumar, M. *et al.* (2014) Tobacco rattle virus-based virus-induced gene silencing in *Nicotiana benthamiana*. *Nat. Protoc.* 9, 1549–1562 29
5. Ratcliff, F. *et al.* (2001) Technical advance. Tobacco rattle virus as a vector for analysis of gene function by silencing. *Plant J.* 25, 237–245 30
6. Dommès, A.B. *et al.* (2019) Virus-induced gene silencing: empowering genetics in non-model organisms. *J. Exp. Bot.* 70, 757–770 31
7. Bond, D.M. *et al.* (2015) Epigenetic transitions leading to heritable, RNA-mediated *de novo* silencing in *Arabidopsis thaliana*. *Proc. Natl Acad. Sci. U. S. A.* 112, 917–922 32
8. Liu, E. *et al.* (2008) Optimized cDNA libraries for virus-induced gene silencing (VIGS) using tobacco rattle virus. *Plant Methods* 4, 5 33
9. Pacak, A. *et al.* (2010) Investigations of barley stripe mosaic virus as a gene silencing vector in barley roots and in *Brachypodium distachyon* and oat. *Plant Methods* 6, 26 34
10. Sasaki, S. *et al.* (2010) Efficient virus-induced gene silencing in apple, pear and Japanese pear using apple latent spherical virus vectors. *Plant Methods* 7, 15 35